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Enzymatic Synthesis of Capsaicin Analogs with Liver Acetone Powder

Kenji Kobata, Koichi Yoshikawa, Masahiro Kohashi,
and Tatsuo Watanabe

School of Food and Nutritional Sciences, University of Shizuoka, 52-1 Yada,
Shizuoka-shi, Shizuoka 422, Japan

Abstract: The enzymatic synthesis of capsaicin analogs with saturated or unsaturated acyl moieties has been achieved by using liver acetone powder as a catalyst. Copyright © 1996 Elsevier Science Ltd

Capsaicin (CAP) is a major pungent principle of capsicum fruits and has many bioactivities.^{1,2} However, the strong pungency and nociceptive activity of CAP seem to inhibit its usage as a food additive or a drug. Several non-pungent CAP analogs (including **4** and **5**) cause adrenal catecholamine secretion.³ A (*Z*)-9-octadecenoyl vanillylamide named olvanil (**6**) is known to be an antinociceptive and antiinflammatory analgesic without initial nociceptive reaction.⁴⁻⁶

To establish the facile supply of CAP analogs, we studied an enzymatic synthesis proceeding under mild conditions and without poisonous agents.

Applications of crude enzyme have gained importance in organic transformations because they are a cheap and easily available biocatalyst. Basavaiah et al. reported enzymatic hydrolysis using bovine liver acetone powder as a catalyst.^{7,8} On the other hand, Park et al. recently reported that the CAP hydrolase was extracted from rat liver microsome.⁹ We therefore used liver acetone powder as an enzyme source.

Vanillylamine hydrochloride could be condensed with a fatty acid methyl ester by using liver acetone powder. This reaction proceeded in a two-phase system in which an oil phase itself consists of the fatty acid methyl ester used as a substrate.

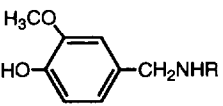
Reactive conditions were examined with methyl tetradecanoate (methyl myristate: C_{14:0}-Me) as a model substrate, and the yields were analyzed by HPLC method.¹⁰

The CAP analog with a tetradecanoyl moiety (myristoyl: C_{14:0}, **4**) was obtained under the following conditions. Ten μmol of vanillylamine hydrochloride (1.9 mg) and 1000 μmol of methyl tetradecanoate (275 μl) were dissolved in 500 μl of 10 mM borate buffer, pH 9.0, and 10 μmol of diisopropylethylamine (1.7 μl) was added. Further, 10 mg of chicken liver acetone powder (Sigma Chem. Co., St. Louis, MO) was added, and the solution was stirred with a magnetic stirrer at 37 °C in a water bath. The yield after the 24 h reaction

under this condition was 14.8%. No product was obtained when deactivated enzyme, or no enzyme, was used. The reaction was then performed on a twenty times larger scale, and the structure of the product was confirmed by $^1\text{H-NMR}$ method.¹¹

Various CAP analogs, octanoyl ($\text{C}_{8:0}$, **1**), decanoyl (caproyl: $\text{C}_{10:0}$, **2**), dodecanoyl (lauroyl: $\text{C}_{12:0}$, **3**), hexadecanoyl (palmitoyl: $\text{C}_{16:0}$, **5**) and (*Z*)-9-octadecenoyl (oleoyl: $\text{C}_{18:1}$, **6**) vanillylamides could be synthesized under the same conditions as mentioned above. The yields of each analog were analyzed by HPLC method and are shown in Table 1.

Table 1. The yields of various CAP analogs (1-6) synthesized by enzymatic synthesis

Capsaicin: R= $\text{CO}(\text{CH}_2)_4\text{CH}=\text{CHCH}(\text{CH}_3)_2$	R	Reaction time and yield (%)	
		24 h	48 h
	1 $\text{CO}(\text{CH}_2)_6\text{CH}_3$	7.4	8.3
	2 $\text{CO}(\text{CH}_2)_8\text{CH}_3$	25.0	27.6
	3 $\text{CO}(\text{CH}_2)_{10}\text{CH}_3$	21.5	17.5
	4 $\text{CO}(\text{CH}_2)_{12}\text{CH}_3$	14.8	17.2
	5 $\text{CO}(\text{CH}_2)_{14}\text{CH}_3$	9.3	12.9
	6 $\text{CO}(\text{CH}_2)_7\text{CH}=\text{CH}(\text{CH}_2)_7\text{CH}_3$	8.4	13.4

In summary, CAP analogs with acyl moieties of various chain lengths could be synthesized by the use of a cheap and easily available biocatalyst, *i.e.*, liver acetone powder.

References and Notes

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10. Preparation of the HPLC sample: after centrifuging of the reaction mixture at 3000 rpm for 5 min, one μl of the upper layer was injected. HPLC conditions: column, Wakopak 5C18 AR, 48 x 150 mm (Wako Pure Chem. Ind., Osaka, Japan); eluent, methanol; flow rate, 0.5 ml/min; detection, UV 280 nm.
11. $^1\text{H-NMR}$ spectrum of **4**: δ (CDCl_3): 6.81 (1H, d, $J=8.4$ Hz), 6.76 (1H, s), 6.71 (1H, d, $J=8.4$ Hz), 5.60 (1H, bt), 5.56 (1H, s), 4.31 (2H, d, $J=6.0$ Hz), 3.83 (3H, s), 2.14 (2H, t, $J=7.6$ Hz), 1.60 (2H, quint, $J=7.6$ Hz), 1.20 (20H, s), 0.83 (3H, t, $J=7.6$ Hz).

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